



REVIEW

Expert Consensus on the Use of Prognostic Gene Expression Profiling Tests for the Management of Cutaneous Melanoma: Consensus from the Skin Cancer Prevention Working Group

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ABSTRACT

Background: Prognostic assessment of cutaneous melanoma relies on historical, clinicopathological, and phenotypic risk factors according to American Joint Committee on

Cancer(AJCC) and National Comprehensive Cancer Network (NCCN) guidelines but may not account for a patient's individual additional genetic risk factors.

Objective: To review the available literature regarding commercially available gene expression profile (GEP) tests and their use in the management of cutaneous melanoma.

Methods: A literature search was conducted for original, English-language studies or meta-analyses published between 2010 and 2021 on commercially available GEP tests in cutaneous melanoma prognosis, clinical decision-making regarding sentinel lymph node biopsy, and real-world efficacy. After the literature review, the Skin Cancer Prevention Working Group, an expert panel of dermatologists with specialized training in melanoma and non-melanoma skin cancer diagnosis and management, utilized a modified Delphi technique to develop consensus statements regarding prognostic gene expression profile tests. Statements were only adopted with a supermajority vote of > 80%.

Results: The initial search identified 1064 studies/meta-analyses that met the search criteria. Of these, we included 21 original articles and meta-analyses that studied the 31-GEP test (DecisionDx-Melanoma; Castle Biosciences, Inc.), five original articles that studied the 11-GEP test (Melagenix; NeraCare GmbH), and four original articles that studied the 8-GEP test with clinicopathological factors (Merlin; 8-GEP + CP; SkylineDx B.V.) in this review. Six

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statements received supermajority approval and were adopted by the panel.

Conclusion: GEP tests provide additional, reproducible information for dermatologists to consider within the larger framework of the eighth edition of the AJCC and NCCN cutaneous melanoma guidelines when counseling regarding prognosis and when considering a sentinel lymph node biopsy.

Keywords: Cutaneous melanoma; Gene-expression profile; Consensus; American joint committee on cancer 8th edition; Prognostic staging; Risk; Prediction; Sentinel lymph node; Sentinel lymph node biopsy

Key Summary Points

Cutaneous melanoma is a growing public health concern, with annual incidence increasing by 3% per year and 7180 individuals expected to die from cutaneous melanoma in 2021 alone.

The current AJCC8 and NCCN guidelines utilize historical, pathological, and phenotypic risk factors to determine melanoma prognosis, but these do not account for genomic expression and may not optimize melanoma prognostic assessment.

Gene expression profile (GEP) tests are validated, reproducible, and consistent across studies.

Studies have demonstrated that integrating GEPs into AJCC8 and NCCN models can improve prognosis and clinical decision-making regarding sentinel lymph node biopsies.

Incorporating GEP testing into real-world clinical management has positively impacted patient outcomes.

INTRODUCTION

Cutaneous melanoma (CM) is the sixth most common malignancy in the USA [1–4]. It is estimated that one in 27 men and one of 40 women will receive a melanoma diagnosis in their lifetime [1–4]. Despite earlier diagnoses and management leading to fewer annual CM-related deaths, 7180 Americans are expected to die from CM in 2021, accounting for over \$1.5 billion in annual healthcare spending [2, 5, 21].

Patients with CM are primarily staged according to the American Joint Committee on Cancer (8th edition [AJCC8]) criteria, including Breslow thickness, ulceration, sentinel lymph node (SLN) status, and presence of distant metastasis [6]. Patients without lymphatic spread or distant metastasis (stages I–II) typically have a better long-term prognosis or melanoma-specific survival (MSS) compared to those with SLN involvement (stage III) or distant metastasis (stage IV) [6, 7]. Based on these factors, National Comprehensive Cancer Network (NCCN) guidelines recommend increasing degrees of clinical management [8]. However, “low-risk” stage I–IIA CM still incurs morbidity and mortality, with a 5-year MSS of 99% for stage IA CM, 97% for stage IB CM, and 94% for stage IIA CM per AJCC8 model [6, 7].

Studies have identified additional factors outside of AJCC staging with varying degrees of prognostic utility, including the number of mitoses/mm², tumor regression, tumor-infiltrating lymphocytes, lymphovascular invasion, tumor location, uncertain microstaging, and patient age [6–9]. Gene expression profile (GEP) tests were developed to gain insight into the tumor molecular biology to assist in prognostic assessment [9, 10]. Despite advancements in GEP technology and the increasing common use of GEP testing across other notable malignancies, including breast, prostate, lung, and colorectal cancer, controversy regarding their clinical implementation and validity in CM prognosis persists [11–16].

The purpose of this study was to review the available literature regarding the validity, accuracy, efficacy, and utility of commercially

available prognostic GEP tests for CM and provide insight into the nuances of current controversies and real-world applications of GEP testing.

METHODS

Literature Search

A MEDLINE search was performed using the keywords “cutaneous melanoma,” “primary melanoma,” “gene expression profile,” “prognosis,” “risk,” and “sentinel lymph node biopsy” and the Boolean terms “AND” and “OR” for full-length, original research, English-language articles and meta-analyses published between 2010 and 2021. Articles were screened, appraised, and selected based on Oxford Center for Evidence-based Medicine criteria for relevance investigating GEP use in augmenting CM prognosis, sentinel lymph node biopsy (SLNBx), and real-world clinical decision-making [16]. References from selected articles were also reviewed for relevant articles not found in the initial search. Final articles were distributed to members of the consensus panel for individual review.

Consensus Development Process

An eight-person consensus panel of dermatologists representing the Skin Cancer Prevention Working Group (SCPWG), comprised of physicians with additional specialized training in managing and diagnosing melanoma and non-melanoma skin cancers, convened on 28 October 2021 and 28 December 2021 to discuss issues surrounding the clinical implementation and appropriate use of GEP testing. Consensus statements were constructed based on the review and discussion of the selected articles.

A modified Delphi technique was used to achieve consensus among panel members [18]. The modified Delphi technique has been previously employed in the development of dermatologic expert panel consensus recommendations [18–20]. This technique utilizes serial rounds of real-time voting with a required

supermajority (> 80%) to adopt a proposed statement. Statements undergo additional discussion, modification, and additional rounds of voting if supermajority approval is not achieved.

Ethics

The article is based on previously conducted studies and does not contain any new studies with human participants or animals performed by any of the authors.

RESULTS

The initial search identified 1064 studies/meta-analyses that met the search criteria. In reviewing the available literature of prognostic GEP tests, the panel found 21 original articles and meta-analyses that studied the 31-GEP test (DecisionDx-Melanoma; Castle Biosciences, Inc., Friendswood, TX, USA) [14, 31–33, 39–49, 59–67], five original articles that studied the 11-GEP test (Melagenix; NeraCare GmbH, Bönen, Germany) [14, 34–36, 50], and four original articles that studied the 8-GEP + CP test (Merlin; SkylineDx B.V., Rotterdam, the Netherlands) [37, 38, 51, 57].

The expert consensus panel developed six statements that all received supermajority approval using the modified Delphi technique (Table 1).

Cutaneous Melanoma is a Growing Public Health Concern

Panel members note that while not a new public health concern, the overall impact CM has on population health continues to grow at a rate of approximately 3% annually [22–26]. While there is contention regarding potential overdiagnosis, especially of thinner melanomas, recent epidemiological studies suggest that improvements in screening and diagnostic techniques have decreased the incidence of thicker melanomas [22–26]. Given the increasing CM incidence, the panel further emphasizes the importance of appropriate and optimized

Table 1 Consensus statements arrived at by the expert consensus panel using the modified Delphi technique

Statement	Consensus (n)
Cutaneous melanoma is a growing public health concern	8/8
The current AJCC8 and NCCN guidelines utilize historical, pathologic, and phenotypic risk factors to determine melanoma prognosis	8/8
The AJCC8 and NCCN prognostic model, which does not account for genomic expression, may not optimize melanoma prognostic assessment	7/8
Gene expression profile (GEP) tests are validated, reproducible, and consistent across studies	8/8
Integrating GEPs into AJCC8 and NCCN models can improve prognostic accuracy	8/8
Prognostic GEP tests can inform clinical decision making regarding sentinel lymph node biopsies	8/8
Incorporating GEPs into real-world clinical management has positively impacted patient outcomes	8/8

AJCC8 American Joint Committee on Cancer (8th edition), *GEP* gene expression profile, *NCCN* National Comprehensive Cancer Network

resource allocation to best identify high-risk patients and populations that would benefit from increased clinical scrutiny and management.

The Current AJCC8 and NCCN Guidelines Utilize Historical, Pathological, and Phenotypic Risk Factors to Determine Melanoma Prognosis

The AJCC8 and NCCN guidelines provide a prognostic assessment of MSS based on clinicopathological CM stages [6–9]. AJCC8

guidelines focus on permutations of three factors: thickness (T), regional nodal and non-nodal metastasis (N), and distant metastases (M) (Table 2) and also include histological factors (e.g., ulceration) and, in advanced disease, serum lactate dehydrogenase levels [6–9]. Higher AJCC8 stages carry a worse 5-year MSS and an implied higher risk of local recurrence and distant metastasis (after complete surgical excision) [6, 7]. NCCN guidelines also include number of mitoses/mm², presence of BRAF mutations, and additional histopathological factors (e.g., desmoplastic subtypes) to guide clinical decision-making based on inherent and potential risk [8].

The AJCC8 and NCCN Prognostic Model, Which Does not Account for Genomic Expression, may not Optimize Melanoma Prognostic Assessment

The AJCC8 and NCCN guidelines account for a large range of outcomes even among “lower-risk” CM stages, with an MSS of 99% for stage IA CM and 97% for stage IB CM, to 94% and 82% for stage IIA and stage IIC CM, respectively [6–8, 27]. There are also instances wherein lower AJCC8 CM stages carry a higher mortality than higher AJCC8 CM stages; for example, patients with stage IIIA disease have a 5-year MSS of 93% compared to those with a “lower” stage IIC disease, who have a 5-year MSS of 82% [6, 7].

An important clinical consideration is that while thin CM (≤ 1.0 mm Breslow depth) often have a good MSS, by sheer volume, stage I CM account for approximately 80% of all melanomas diagnosed. As a result, these thinner, “lower-risk” CM account for approximately 26% of all melanoma-specific mortality (MSM) [9, 27, 28]. Therefore, although thin CM may carry a better MSS, there may be a currently unidentified subset of patients with thin CM that have inherently increased morbidity (e.g., worse 5-year MSS, increased risk of recurrence, and potential metastasis). Additional factors (e.g., genetic mutations and tumor biology) beyond those included in AJCC8 staging likely contribute to the observed 5-year MSS overlap during the initial staging process. [29, 30].

Table 2 The American Joint Committee on Cancer (8th edition) melanoma clinicopathological staging and associated 5-year melanoma specific survival

TNM staging system		Description	5-year MSS (%)	
Overall stage	Subcategory			
T		Thickness	Ulceration	
T1	T1a	< 0.8 mm, no ulceration	No	99
	T1b	≤ 0.8 mm, with ulceration	Yes	99
T2	T2a	> 1–2 mm, no ulceration	No	96
	T2b	> 1–2 mm, with ulceration	Yes	93
T3	T3a	> 2–4 mm, no ulceration	No	94
	T3b	> 2–4 mm, with ulceration	Yes	86
T4	T4a	> 4 mm, no ulceration	No	90
	T4b	> 4 mm, with ulceration	Yes	82
N		Number of nodes	In-transit or (micro)satellite metastases	
N1	N1a	1 clinically occult lymph node, detectable by SLNBx	No	84
	N1b	1 clinically detectable lymph node	No	76
	N1c	No regional lymph node disease	Yes	81
N2	N2a	2–3 clinically occult lymph node, detectable by SLNBx	No	79
	N2b	2–3 nodes, ≥ 1 clinically detectable node	No	71
	N2c	1 clinically occult or clinically detected	Yes	69
N3	N3a	≥ 4 clinically occult lymph node, detectable by SLNBx	No	60
	N3b	≥ 4 with ≥ 1 clinically detected or presence of matted nodes	No	64
	N3c	≥ 2 clinically occult/detectable nodes and/or presence of matted nodes	Yes	52
M1		Anatomic site	LDH elevated	
M1a	M1a(0)	Distant metastasis to skin, soft tissue, and/or nonregional lymph node	No	–
	M1a(1)		Yes	–
M1b	M1b(0)	Distant metastasis to lung ± M1a sites	No	–
	M1b(1)		Yes	–
M1c	M1c(0)	Distant metastasis to non-CNS visceral sites ± M1a or M1b sites	No	–
	M1c(1)		Yes	–
M1d	M1d(0)	Distant metastasis to CNS ± M1a-M1c sites	No	–
	M1d(1)		Yes	–

Adapted from Gershenwald et al. [7]; “–” indicates data are not available in Gershenwald et al. [7]

CNS Central nervous system, LDH lactate dehydrogenase, MSS melanoma-specific survival, SLNBx sentinel lymph node biopsy

Identifying these higher-risk thin CM may provide an opportunity to identify patients who may benefit from additional clinical scrutiny, individualized follow-up intervals, multispecialty referrals, radiographic monitoring, and adjuvant therapy.

GEP tests are Validated, Reproducible, and Consistent Across Studies

GEP tests use tumor tissue obtained from the initial biopsy to assess how different genes are regulated, thereby providing additional objective data for clinical decision-making [10, 28]. Several GEP tests have been developed and validated to supplement prognosis in stage I–III CM: the 31-GEP test [31–33], the 11-GEP test [34–36], and the 8-GEP + CP test [37, 38]. The authors note that different genes are assessed in each of these three GEP tests (Table 3). This may be due to the creation of these tests using retrospective methods among different study populations (e.g., 2 German trial sites for the 11-GEP test [35], 3 US-based sites for the 8-GEP + CP test [37], and 6 US-based sites for the 31-GEP test [31]) and/or focusing on different gene functions (e.g., the 11-GEP test focuses on genes associated with lower-risk CM [35], and the 8-GEP + CP test focuses on genes associated with angiogenesis/hypoxia, coagulation, and epithelial-to-mesenchymal transition [37]). Although different genes are assessed between GEP tests, multiple independent studies (prospective) and meta-analyses have demonstrated these tests' internal validity and prognostic consistency. Of note, the authors of this review found that among the original studies for each of these three commercially available GEP tests investigating morbidity (Table 4) and SLNBx status, available literature on the 31-GEP test was fourfold that available on the other two GEP tests, including several meta-analyses and prospective clinical trials with reproducible results, as well as outcomes from real-world clinical management studies [14, 31–33, 39–49, 59–67].

Integrating GEPs into AJCC8 and NCCN Models can Improve Prognostic Accuracy

AJCC8 and NCCN models have material MSS overlap between distinct clinical stages; furthermore, they do not account for all clinically useful prognostic assessments, such as relapse-free or recurrence-free survival (RFS), which are an often used outcome for adjuvant therapy trials, or distant metastasis-free survival (DMFS) [6–9]. Multiple independent studies and meta-analyses have shown that GEP tests can risk-stratify patients to provide more granular interval assessments of RFS, DMFS, and MSM or survival (MSS) [39–51].

31-GEP Test

The prognostic 31-GEP test (DecisionDx-Melanoma; Castle Biosciences, Inc.) is a Medicare-reimbursable test that uses 28-gene targets and three control genes to assess tumor biology. Retrospective and prospective cohort studies determined that stage I–III CM designated as high risk (e.g. 31-GEP class 2) carried a significantly worse 5-year RFS [33, 39–42], 3-year RFS [44–46], 5-year MSS [33, 39–42], 5-year DMFS [39], 3-year DMFS [44–46], 5-year MSS [39–42] and metastasis-free survival (MFS) [39–42] than similarly staged GEP class 1 patients (Table 4). Prospective studies have also demonstrated that the 31-GEP test is a significant, independent predictor of RFS and DMFS for the “low-risk” AJCC8 stage I–IIA CM [47].

Data from prospective cohorts and the EXPAND and INTEGRATE clinical trials found the 31-GEP test could stratify AJCC8 prognostic models further [43]. Among patients considered at lower risk by AJCC staging (e.g., stage I–IIA CM), those with a 31-GEP class 2 result had significantly lower 3-year survival than those with a class 1 result for RFS (83 vs. 97%; $p < 0.001$), DMFS (87 vs. 99%; $p < 0.001$), and overall survival (OS) (90 vs. 98%; $P = 0.01$) [37]. Similar trends were noted for patients considered to be higher risk AJCC staging (e.g., stage IIB–III CM) for 3-year RFS (class 2: 52% vs. class

Table 3 Genes assessed in commercially available gene expression profile tests for cutaneous melanoma

31-GEP test		11-GEP test		8-GEP + CP test
Upregulated	Downregulated	Risk	Protective	
Secreted phosphoprotein 1	BRCA1-associated protein	Kelch-like family member 41	Keratin 9	Melanoma antigen recognized by T cells 1
Keratin 6B	Matrix Gla protein	Esophageal cancer-related gene 2	Dermicidin	Growth differentiation factor 15
Eukaryotic translation initiation factor 1B	Chemokine (CXC motif) ligand 14	Hairy and enhancer of split 6	Prolactin-induced protein	CXCL8
	S100 calcium-binding protein A8		Secretoglobin family 1D member 2	Lysyl oxidase homolog 4
	B-cell translocation gene 1, antiproliferative		Secretoglobin family 2A member 2	TGF-β receptor type 1
	Sin3A-associated protein, 130 kDa		Collagen alpha6	Integrin- β3
	Gap junction protein, alpha 1, 43 kDa		Guanylate binding protein 4	Tissue-type plasminogen activator
	Inhibitor of DNA binding 2, dominant negative helix-loop-helix protein		Mucin 7	Glia-derived nexin
	S100 calcium-binding protein A9			
	Cellular retinoic acid binding protein 2			
	Keratin 14			
	Roundabout, axon guidance receptor, homolog 1 (Drosophila)			
	RNA-binding motif protein 23			
	Tumor-associated calcium signal transducer 2			
	Desmocollin 1			
	Small proline-rich protein 1B			
	Tripartite motif containing 29			
	Aquaporin 3 (Gill blood group)			

Table 3 continued

31-GEP test		11-GEP test		8-GEP + CP test
Upregulated	Downregulated	Risk	Protective	
	Tyrosinase-related protein 1			
	Periplakin			
	Leukotriene A4 hydrolase			
	Cystatin E/M			

1: 79%; $p = 0.02$), DMFS (class 2: 74% vs. class 1: 79%; $P = 0.40$), and OS (class 2: 74% vs. class 1: 91%; $p = 0.02$) [43].

In addition, studies found increased precision using 31-GEP subclasses, with the class 2B designation carrying the worst 3-year RFS (60%), DMFS (78%), and OS (74%) compared to class 1A denoting lowest risk [43]. Importantly, clinical trial data have found that a stage I-IIA class 2B CM and stage IIB-III CM had similar rates of distant metastasis (21 vs. 24%) and deaths (29 vs. 22%) [43].

In multivariate meta-analyses [48, 49], retrospective studies [32, 33, 39, 41, 42], and prospective studies [43–47], the 31-GEP test has consistently been found to be a significant predictor with a strong negative predictive value (NPV) for RFS (NPV 94%) [43], DMFS (NPV 97%) [43], and OS (NPV 97%) [43] for class 1 patients, independent of AJCC8 prognostic criteria, including Breslow depth, ulceration, and SLN status.

11-GEP Test

The 11-GEP (Melagenix; NeraCare GmbH) uses eight prognostic targets and three reference genes to assess CM via a continuous scoring system in which “0” delineates “low-risk” (i.e., ≤ 0) versus “high-risk” (> 0) [44, 45]. A retrospective study of 291 stage I-III CMs and prospective study of 245 stage II CMs found that high-risk patients identified with the 11-GEP test have a significantly worse 5-year disease-free survival (DFS) ($p < 0.001$) and 5-year MSS ($p = 0.001$) than their low-risk counterparts (Table 4) [36]. In a prospective study of 245 stage II CM, high-risk patients had significantly worse 5-year RFS ($p = 0.009$), DMFS ($p = 0.005$),

MSS ($p = 0.018$), and 10-year MSS ($p = 0.018$) (Table 4) [50]. In both studies, multivariate analysis found the continuous ($p < 0.0068$) [30] and binary stratification ($p = 0.018$) [50] 11-GEP to be a significant prognostic factor for MSS, independent of age or Breslow thickness [36, 50]. Of note, initial data also suggest a potential synergy of the 11-GEP with AJCC8 staging [35].

8-GEP + CP Test

The 8-GEP + CP test (Merlin, SkylineDx B.V.) is a logistic regression model comprised of eight genes and two clinicopathological factors initially designed and validated to predict SLN status [37, 38]. The model was then retrained for prognostic assessment in stage I-IIA CM [51]. In a retrospective study, the 8-GEP + CP test was found to be an independent predictor of 5-year RFS (stage I-III CM, $p < 0.001$; stage I-IIA, $p = 0.006$) and 5-year DMFS (stage I-III CM, $p = 0.001$; stage I-IIA CM, $p = 0.025$), but not of 5-year MSS, after accounting for age and Breslow thickness [51]. The 8-GEP + CP separated patients into high- and low-risk prognostic categories by 5-year RFS and DMFS with additional stratification for patients with SLN status and among patients with AJCC8 stages I-IIA (Table 4) [51]. The 8-GEP + CP did not significantly stratify/differentiate MSS between risk classes [51].

Prognostic GEP Tests can Augment Clinical Decision-Making Regarding SLN Biopsies

According to NCCN guidelines, SLNBx is not recommended for patients with $< 5\%$

Table 4 Prognostic end-points for original studies of commercially available gene-expression profile tests

GEP test	Study	Design	CM AJCC8 stage (n)	GEP Risk	3-Year RFS (%)	p value	3-Year DMFS (%)	p value	3-Year MSS (%)	p value
31-GEP	Keller et al. 2019 [44]	Prospective	Stage I-III (159)	Class 1	96	< 0.0001	99	< 0.0001	- ^e	- ^e
				Class 2	47		64			
	Hsueh et al. 2021 [43]	Prospective (clinical trial)	Stage I-III (323)	Class 1	95	0.02	97	0.4	97 ^f	0.02
				Class 2	66		79		81 ^f	
				Class 1	97	< 0.0001	99	< 0.0001	98 ^f	0.01
				Class 2	83		87		90 ^f	
31-GEP	Ferris et al. 2017 [41]	Retrospective	CM AJCC8 stage Stage I-IIA (135)	GEP risk		p value	5-Year DMFS (%)	p value	5-Year MSS (%)	p value
				Class 1	95	< 0.05	96	< 0.05	96 ^f	< 0.05
	Gastrman et al. 2019 [42]	Retrospective	Stage I-III (157)	Class 2	62		76		71 ^f	
				Class 1	75	< 0.05	92	< 0.05	83 ^f	< 0.05
	Zager, et al. 2018 [39]	Retrospective	Stage I (264)	Class 2	17		39		44 ^f	
				Class 1A	80	< 0.0001	83	< 0.0001	98	< 0.0001
				Class 1B	74		74		90	
				Class 2A	46		50		84	
				Class 2B	25		33		61	
				Class 1	96	0.01 ^d	97	0.085 ^d	99	0.37 ^d
Greenhaw et al. 2018 [40]	Retrospective	Stage II (93)	Class 2	85		90		97		
			Class 1A	98	< 0.001 ^d	98	0.05 ^d	100	< 0.01 ^d	
			Class 2B	73		87		93		
			Class 1	74	0.043 ^d	90	0.004 ^d	100	0.022 ^d	
			Class 2	55		63		87		
			Class 1A	77	0.13 ^d	95	< 0.001 ^d	100	0.13 ^d	
			Class 2B	50		57		82		
			Class 1	72	0.015 ^d	80	0.019 ^d	100	0.009 ^d	
			Class 2	51		54		67		
			Class 1	93 ^b	< 0.00001	- ^e	- ^e	99	0.00003	
Class 2	69 ^b		79		79					

Table 4 continued

GEP test	Study	Design	CM AJCC8 stage (n)	GEP Risk	3-Year RFS (%)	p value	3-Year DMFS (%)	p value	3-YearMSS (%)	p value
11-GEP	Gambichler et al. 2020 [36]	Retrospective	Stage I-III (291)	≤ 0	96 ^c	< 0.0001	- ^e	- ^e	99	0.001
				> 0	78 ^c				88	
	Almaral et al. 2019 [50]	Prospective	Stage II (245)	≤ 0	76 ^c	0.009	89	0.005	92	0.018
				> 0	58 ^c		70		82	
8-GEP + CP	Eggermont et al. 2020 [51]	Retrospective	Stage I-III (837)	Low	87	< 0.001	92	0.001	96	0.064
				High	62		72		88	
			Stage I-III, SLNBx(-)	Low	89	< 0.001	94	0.002	96	0.152
			(637) ^a	High	70		78		89	
			Stage I-IIA	Low	89	0.006	94	0.025	97	0.123
			(580) ^a	High	74		80		91	

CM Cutaneous melanoma, DMFS distant metastasis-free survival, RFS recurrence-free survival,

^aSubset analysis of study sample

^bMetastasis-free survival

^cDisease-free survival

^dAdjusted p value

^eData not available from original paper (-)

^fOverall survival

likelihood of a positive node (e.g., T1a CM without additional risk factors), should be discussed with patients with a 5–10% likelihood of positivity (e.g., T1b or T1a with risk factors, including transected base, age < 40, or > 1 mitosis/mm²), and should be offered to patients with > 10% likelihood of SLN positivity (e.g., clinicopathological stage IB or higher) [8]. The panel notes the importance of holistically discussing the risks and benefits of SLNBx, including low detection rates (approx. 12–20%) [52], false-positive and false-negative rates, approximately 10.1% postoperative morbidity (including wound dehiscence, hematoma/seroma formation, and surgical site infection), as well as financial impact, with one study estimating a cost of \$47,906 to detect one positive SLN [52–54].

31-GEP Test

In a study of prospectively tested patients with T1/T2 CM, a 31-GEP class 1A designation was found to predict a < 5% likelihood of SLN positivity among patients aged ≥ 65 years [46]. Furthermore, the 31-GEP test could further stratify prognosis according to risk when subclass designations are combined (e.g., class 1A or class 2B) with SLN status [46]. Among patients with T1/T2 SLN-negative CM aged ≥ 55 years, those with a 31-GEP class 2B result had significantly worse 5-year RFS (66.7 vs. 91.4%; $p < 0.05$), DMFS (76.2 vs. 93.4%; $p < 0.05$), and MSS (85.4 vs. 99.3%; $p < 0.05$) than patients with a class 1A result [46]. Prognostic differences were starker when patients with T1/T2 tumors, aged ≥ 55 years, who received a class 2B 31-GEP result and were SLN positive were compared to their class 1A counterparts in terms of 5-year RFS (19.1 vs. 91.4%; $p < 0.001$), DMFS (32.1 vs. 93.4%; $p < 0.001$), OS (18.5 vs. 96.3%; $p < 0.001$) and MSS (55.0 vs. 99.3%; $p < 0.001$) [46]. An additional study published after the SCPWG consensus meeting found an integrated 31-GEP (i31-GEP) test using a continuous score and traditional clinicopathological factors had a 98% NPV, was able to accurately identify up to 27% of 1674 patients with T1-4 CM, and was able to reclassify an additional 63% of patients with an intermediate probability of SLN positivity (5–10%) [55]. The

additive prognostic ability and increased sensitivity and specificity of the 31-GEP and SLN status were also demonstrated in additional meta-analyses and clinical trials [43, 49].

8-GEP + CP Test

In development, the 8-GEP + CP test, which incorporates the expression of several tumor cell-adhesion genes found to be associated with SLN metastasis (e.g., integrin-β3 and tissue-type plasminogen activator), was noted to have a high NPV capable of identifying tumors (T1-T3) at a low risk of SLN metastasis [37, 56]. In two separate retrospective cohorts, the 8-GEP + CP had an NPV point estimate of 93.3–100% for T1 CM (T1a $n = 8$; T1b $n = 77$), 89.3%–93.3% for T2 CM, and 75.0%–100% for T3 CM [38, 57]. From a cohort of 208 patients, NPV point estimates were 92.9% for T1 CM and 100% for T2 and T3 CM for those aged ≥ 65 years [38]. Retrospective studies on the 8-GEP + CP also report an “SLNBx reduction rate” (or proportion of patients the 8-GEP + CP assessment alone would designate as “low-risk”) of 60.8% among patients with T1 CM, 24.1% for those with T2 CM, and 2.5% for those with T3 CM. Of note, this metric does not account for the error rate, noted to be 2–3% depending on the CM stage [38].

11-GEP Test

There were no relevant articles found within the literature search regarding the 11-GEP and SLNBx prognosis/prediction.

Incorporating GEPs into Real-World Clinical Management has Positively Impacted Patient Outcomes

In 2020–2021, a reported 27,051 patients were clinically tested using the 31-GEP [58], with a 2020 study estimating that 45.2% of dermatologists [59] ordered the test within the 2020 calendar year. More importantly, studies suggest that integrating 31-GEP results into traditional clinical management has positively impacted patient outcomes [27, 59–65].

31-GEP Test

One multicenter study found that in patients with stage I-II CM who were cared for by dermatologists and surgical/medical oncologists, 49% had their management directly altered by 31-GEP class results [60]; specifically, 91% of decreases in management intensity occurred in patients with a class 1 result [60]. In comparison, 72% of increased management intensity occurred in patients with a class 2 result, with significant management differences for patients with higher-risk 31-GEP classes regarding frequency of follow-up visits ($p < 0.001$) [52], imaging ($p < 0.001$) [52], and laboratory testing ($p = 0.04$) [60].

A single-center multidisciplinary prospective study also found a significantly increased likelihood of patients with a class 2 designation following up with surgical oncology and receiving a recommendation for adjuvant trial (stage I, 100 vs. 18%, $p < 0.001$; stage II, 64 vs. 36%, $p < 0.05$) versus patients with class 1 CM [61].

Additional studies have also found GEP class results affected the frequency of physical exams ($p < 0.0001$) [54] and of referrals ($p < 0.0001$) [62], with significantly more physicians altering management and opting to refer CM patients with a 0.7-mm Breslow depth if they also had a GEP class 2 result ($p < 0.05$) [63]. Furthermore, approximately 65% of class 1 results lead to surveillance intensity similar to AJCC8 stage I-IIA, while 98% of class 2 results lead to increased scrutiny, similar to patients with AJCC8 stage IIB-IV CM [64]. Overall, studies have found that 31-GEP class results had the potential to positively influence management decisions among physicians [63–65] and non-physician providers [66], consistent with the augmented risk stratification provided by GEP testing.

11-GEP/8-GEP + CP Test

No relevant articles were found in the literature search on the real-world use of the 11-GEP or 8-GEP + CP.

DISCUSSION

The current AJCC8 [6, 7] and NCCN guidelines [8] provide a framework for a population-based approach for the management of CM. However, while these guidelines do provide a framework for many CM presentations, they may not account for all potential clinically relevant risk factors for an individual patient. As such, the NCCN guidelines recommend that a patient's individual risk of disease recurrence drive clinical decision-making [8]. To that end, it is important that dermatologists have all the tools available to robustly risk-stratify CM patients to provide adequate follow-up, management, and therapy.

There is a large and growing amount of literature demonstrating the validity, efficacy, and utility of GEP tests in the diagnosis and management of CM [10, 12, 13, 27, 31–51, 57, 59–67]. In prognostic assessments of morbidity and mortality, GEP tests may provide additional information to current AJCC8 prognostic models for MSS. [36, 39–42, 50]. Studies also suggest that GEP tests may provide more nuanced information to assist dermatologists in managing CM with regards to SLNBx in conjunction with current NCCN guidelines [37, 38, 43, 46, 49, 54] and potentially reduce unnecessary testing, procedures, and annual healthcare expenditures [28, 60–66]. While meta-analyses are not yet available for all prognostic CM GEP tests, two recent separate studies [49, 67] found similar and robust evidence for the 31-GEP test, which was also supported by clinical recommendations (including A-strength recommendations for the use of the 31-GEP to guide management of patients with negative SLNBx) from an independent consensus panel [10].

Despite these findings, several critical articles have been published regarding GEP testing [14–16]. While additional appraisal is important, context must be provided. Although randomized control trials (RCTs) are the gold standard for interventions [15, 68], prognostic tests are validated by repeated measurements in large cohorts and by meta-analyses that track consistency across studies (though they can be

retro/proactively applied to cohorts within RCTs as has been performed for GEP tests for breast cancer) [69, 70]. These studies also query the lack of U.S. Food and Drug Administration approval. While this may be necessary for interventional therapies, prognostic tests are validated by repeated large prognostic studies and are performed only at accredited laboratories (i.e., with Clinical Laboratory Improvement Amendments [CLIA] and College of American Pathologists [CAP] certification).

Additional studies should include larger, prospective, novel cohorts to provide a more robust assessment of consistent endpoints: RFS, DMFS, and MSM/MSS. These studies are most essential among thin, “low-risk” CM, with the goal of providing better individualized care. Additional meta-analyses may also be performed to determine the repeated accuracy of these tests within various clinical contexts. GEP tests should also be further refined and integrated into current clinicopathological prognostic models to provide a more nuanced, graded risk assessment for dermatologists and patients, as opposed to the majority binary classifications that are currently widely used. Finally, these additional studies may also include real-world data to determine both physician experience and patient outcome regarding the real-world use of GEP tests.

CONCLUSION

CM poses a substantial public health risk, with approximately 100,000 new cases of invasive CM diagnosed in the USA annually. While current diagnostic and prognostic models for CM management can provide identification and risk stratification of suspicious pigmented lesions, studies have found that the incorporation of GEP tests into current algorithms may provide an objective, non-invasive method to improve the accuracy of risk prediction to inform clinical management decisions and optimize patient care. By studying the molecular underpinnings of CM, dermatologists will ideally be able to reduce unnecessary costs and morbidity associated with CM while providing more individualized care to patients.

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REFERENCES

- Centers for Disease Control and Prevention. Leading cancer Cases and deaths, all races and ethnicities, male and female, 2018. <https://gis.cdc.gov/Cancer/USCS/DataViz.html>. Accessed 2021 May 02.
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer statistics. CA: A Cancer Journal for Clinicians. 2021;71:7–33. <https://doi.org/10.3322/caac.21654>.
- Guy GP Jr, Thomas CC, Thompson T, Watson M, Massetti GM, Richardson LC. Vital signs: melanoma incidence and mortality trends and projections—United States, 1982–2030. MMWR Morb Mortal Wkly Rep. 2015;64(21):591–6.
- National Institutes of Health, National Cancer Institute. Surveillance, epidemiology, and end results program (SEER). Cancer stat facts: melanoma of the skin. <https://seer.cancer.gov/statfacts/html/melan.html>. Accessed 15 May 2021.
- Criscione VD, Weinstock MA. Melanoma thickness trends in the United States, 1988–2006. J Invest Dermatol. 2010;130(3):793–7. <https://doi.org/10.1038/jid.2009.328>.
- Keung EZ, Gershenwald JE. The eighth edition American Joint Committee on Cancer (AJCC) melanoma staging system: implications for melanoma treatment and care. Expert Rev Anticancer Ther. 2018;18(8):775–84. <https://doi.org/10.1080/14737140.2018.1489246>
- Gershenwald JE, Scolyer RA, Hess KR, et al. Melanoma staging: Evidence-based changes in the American Joint Committee on Cancer eighth edition cancer staging manual. CA: A Cancer Journal for Clinicians. 2017;67:472–92.
- National Comprehensive Cancer Network. Melanoma: Cutaneous (version 2.2021). https://www.nccn.org/professionals/physician_gls/pdf/cutaneous_melanoma.pdf. Accessed 17 Jul 2021.
- Bajaj S, Donnelly D, Call M, et al. Melanoma prognosis: accuracy of the American Joint Committee on Cancer Staging Manual eighth edition. J Natl Cancer Inst. 2020;112(9):921–8. <https://doi.org/10.1093/jnci/djaa008>.
- Berman B, Ceilley R, Cockerell C. Appropriate use criteria for the integration of diagnostic and prognostic gene expression profile assays into the management of cutaneous malignant melanoma: an expert panel consensus-based modified Delphi process assessment. SKIN The Journal of Cutaneous Medicine. 2019;3(5):291–306. <https://doi.org/10.25251/skin.3.5.1>.
- Meleth S, Reeder-Hayes K, Ashok M, et al. Technology assessment of molecular pathology testing for the eEstimation of prognosis for common cancers. 2014.
- Rigel DS, Marson JW. Prognostic gene expression profiling in cutaneous melanoma: identifying the knowledge gaps and assessing the clinical benefit. Practice update website. <https://www.practiceupdate.com/content/prognostic-gene-expression-profiling-in-cutaneous-melanoma/104406/65/4/1>. Accessed 20 May 2021.

13. Rigel DS, Ceilley RI, Litchman GH, Cockerell CJ. Gene expression profile testing in skin cancer prognosis: the data is clear – it's time to get on board. *SKIN The Journal of Cutaneous Medicine*. 2020;4 (4):304–8. <https://jofskin.org/index.php/skin/article/view/937>
14. Marchetti MA, Coit DG, Dusza SW, et al. Performance of gene expression profile tests for prognosis in patients with localized cutaneous melanoma: a systematic review and meta-analysis. *JAMA Dermatol*. 2020;156(9):953–62. <https://doi.org/10.1001/jamadermatol.2020.1731>. PMID: 32745161;PMCID:PMC7391179.
15. Grossman D, Okwundu N, Bartlett EK, et al. Prognostic gene expression profiling in cutaneous melanoma: identifying the knowledge gaps and assessing the clinical benefit. *JAMA Dermatol*. 2020;156(9):1004–11. <https://doi.org/10.1001/jamadermatol.2020.1729> (PMID: 32725204).
16. Chan WH, Tsao H. Prognostic gene expression profiling in cutaneous melanoma: identifying the knowledge gaps and assessing the clinical benefit. *JAMA Dermatol*. 2020;156(9):949–51. <https://doi.org/10.1001/jamadermatol.2020.1730>.
17. Oxford Center for Evidence Based Medicine. <http://www.cebm.net/index.aspx?o=5653>. Accessed 18 May 2021.
18. Hsu C-C, Sandford BA. The Delphi technique: making sense of consensus. *Pract Assess Res Eval*. 2007;12:1–8.
19. Richard MA, Barnette T, Rouzaud M, et al. Evidence-based recommendations on the role of dermatologists in the diagnosis and management of psoriatic arthritis: systematic review and expert opinion. *J Eur Acad Dermatol Venereol* 2014;28s5: 3–12.
20. Gottlieb AB, Levin AA, Armstrong AW, et al. The International Dermatology Outcome Measures Group: formation of patient-centered outcome measures in dermatology. *J Am Acad Dermatol*. 2015;72(2):345–8.
21. Lim HW, et al. The burden of skin disease in the United States. *J Am Acad Dermatol*. 2017;76:958–972.e2. <https://doi.org/10.1016/j.jaad.2016.12.043>.
22. Welch HG, Mazer BL, Adamson AS. The rapid rise in cutaneous melanoma diagnoses. *N Engl J Med*. 2021;384(1):72–9. <https://doi.org/10.1056/NEJMs2019760>.
23. Grossman D, Sweeney C, Doherty JA. The rapid rise in cutaneous melanoma diagnoses. *N Engl J Med*. 2021;384(14): e54. <https://doi.org/10.1056/NEJMc2101980>.
24. Goldsmith SM. The rapid rise in cutaneous melanoma diagnoses. *N Engl J Med*. 2021;384(14):e54. <https://doi.org/10.1056/NEJMc2101980>.
25. Lashway SG, Harris RB, Farland LV, O'Rourke MK, Dennis LK. Age and cohort trends of malignant melanoma in the United States. *Cancers*. 2021;13: 3866. <https://doi.org/10.3390/cancers13153866>.
26. Conforti C, Zalaudek I. Epidemiology and risk factors of melanoma: a review. *Dermatol Pract Concept*. 2021;11(Suppl 1):e2021161S. <https://doi.org/10.5826/dpc.11s1a161s>
27. Kwatra SG, Hines H, Semenov YR, Trotter SC, Holland E, Leachman S. A dermatologist's guide to implementation of gene expression profiling in the management of melanoma. *J Clin Aesthet Dermatol*. 2020;13(11 Suppl 1):s3-14.
28. Whiteman DC, Baade PD, Olsen CM. More people die from thin melanomas (<1 mm) than from thick melanomas (>4 mm) in Queensland, Australia. *J Invest Dermatol*. 2015;135(4):1190–3. <https://doi.org/10.1038/jid.2014.452>.
29. Etkorn JR, Sharkey JM, Grunyk JW, Shin TM, Sobanko JF, Miller CJ. Frequency of and risk factors for tumor upstaging after wide local excision of primary cutaneous melanoma. *J Am Acad Dermatol*. 2017;77(2):341–8.
30. Menezes SL, Kelly JW, Wolfe R, Farrugia H, Mar VJ. The increasing use of shave biopsy for diagnosing invasive melanoma in Australia. *Med J Aust*. 2019;211:213–8.
31. Gerami P, Cook RW, Wilkinson J, et al. Development of a prognostic genetic signature to predict the metastatic risk associated with cutaneous melanoma. *Clin Cancer Res*. 2015;21(1):175–83.
32. Gerami P, Cook RW, Russell MC, et al. Gene expression profiling for molecular staging of cutaneous melanoma in patients undergoing sentinel lymph node biopsy. *J Am Acad Dermatol*. 2015;72(5):780-5.e3.
33. Cook RW, Middlebrook B, Wilkinson J, et al. Analytic validity of DecisionDx-Melanoma, a gene expression profile test for determining metastatic risk in melanoma patients. *Diagn Pathol*. 2018;13(1):13.
34. Brunner G, Reitz M, Heinecke A, et al. A nine-gene signature predicting clinical outcome in cutaneous melanoma. *J Cancer Res Clin Oncol*. 2013;139(2): 249–58. <https://doi.org/10.1007/s00432-012-1322-z> (Epub 2012 Oct 9 PMID: 23052696).
35. Brunner g, Heinecke A, Falk TM, et al. A prognostic gene signature expressed in primary cutaneous

- melanoma: synergism with conventional staging. *JNCI Cancer Spectrum*. 2018; 2: pky032. <https://doi.org/10.1093/jncics/pky032>
36. Gambichler T, Tzagoudis K, Kiecker F, et al. Prognostic significance of an 11-gene RNA assay in archival tissue of cutaneous melanoma stage I–III patients. *Eur J Cancer*. 2021;143:11–8. <https://doi.org/10.1016/j.ejca.2020.10.016>.
 37. Bellomo D, Arias-Mejias SM, Ramana C, et al. Model combining tumor molecular and clinicopathologic risk factors predicts sentinel lymph node metastasis in primary cutaneous melanoma. *JCO Precis Oncol*. 2020;4:319–34. <https://doi.org/10.1200/po.19.00206>.
 38. Yousaf A, Tjien-Fooh FJ, Rentroia-Pacheco B, et al. Validation of CP-GEP (Merlin Assay) for predicting sentinel lymph node metastasis in primary cutaneous melanoma patients: A U.S. cohort study. *Int J Dermatol*. 2021;60(7):851–6.
 39. Zager JS, Gastman BR, Leachman S, et al. Performance of a prognostic 31-gene expression profile in an independent cohort of 523 cutaneous melanoma patients. *BMC Cancer*. 2018;18(1):130. <https://doi.org/10.1186/s12885-018-4016-3>.
 40. Greenhaw BN, Zitelli JA, Brodland DG. Estimation of prognosis in invasive cutaneous melanoma: an independent study of the accuracy of a gene expression profile test. *Dermatol Surg*. 2018;44(12):1494–500. <https://doi.org/10.1097/DSS.0000000000001588>.
 41. Ferris LK, Farberg AS, Middlebrook B, et al. Identification of high-risk cutaneous melanoma tumors is improved when combining the online American joint committee on cancer individualized melanoma patient outcome prediction tool with a 31-gene expression profile-based classification. *J Am Acad Dermatol*. 2017;76(5):818–825.e3. <https://doi.org/10.1016/j.jaad.2016.11.051>.
 42. Gastman BR, Zager JS, Messina JL, et al. Performance of a 31-gene expression profile test in cutaneous melanomas of the head and neck. *Head Neck*. 2019;41(4):871–9. <https://doi.org/10.1002/hed.25473>.
 43. Hsueh EC, DeBloom JR, Lee JH, et al. Long-term outcomes in a multicenter, prospective cohort evaluating the prognostic 31-gene expression profile for cutaneous melanoma. *JCO Precis Oncol*. 2021;5:PO.20.00119. <https://doi.org/10.1200/PO.20.00119>.
 44. Keller J, Schwartz TL, Lizalek JM, et al. Prospective validation of the prognostic 31-gene expression profiling test in primary cutaneous melanoma. *Cancer Med*. 2019;8(5):2205–12. <https://doi.org/10.1002/cam4.2128>.
 45. Podlipnik S, Carrera C, Boada A, Richarz NA, et al. Early outcome of a 31-gene expression profile test in 86 AJCC stage IB–II melanoma patients. A prospective multicenter cohort study. *J Eur Acad Dermatol Venereol*. 2019;33(5):857–62. <https://doi.org/10.1111/jdv.15454>.
 46. Vetto JT, Hsueh EC, Gastman BR, et al. Guidance of sentinel lymph node biopsy decisions in patients with T1–T2 melanoma using gene expression profiling. *Future Oncol*. 2019;15(11):1207–17. <https://doi.org/10.2217/fon-2018-0912>.
 47. Arnot SP, Han G, Fortino J, Han D, Fowler G, Vetto JT. Utility of a 31-gene expression profile for predicting outcomes in patients with primary cutaneous melanoma referred for sentinel node biopsy. *Am J Surg*. 2021;221(6):1195–9. <https://doi.org/10.1016/j.amjsurg.2021.03.028>.
 48. Gastman BR, Gerami P, Kurley SJ, Cook RW, Leachman S, Vetto JT. Identification of patients at risk of metastasis using a prognostic 31-gene expression profile in subpopulations of melanoma patients with favorable outcomes by standard criteria. *J Am Acad Dermatol*. 2019;80(1):149–157.e4. <https://doi.org/10.1016/j.jaad.2018.07.028>.
 49. Greenhaw BN, Covington KR, Kurley SJ, et al. Molecular risk prediction in cutaneous melanoma: A meta-analysis of the 31-gene expression profile prognostic test in 1,479 patients. *J Am Acad Dermatol*. 2020;83:745–53. <https://doi.org/10.1016/j.jaad.2020.03.053>.
 50. Amaral TMS, Hoffmann M-C, Sinnberg T, et al. Clinical validation of a prognostic 11-gene expression profiling score in prospectively collected FPPE tissue of patients with AJCC v8 stage II cutaneous melanoma. *Eur J Cancer*. 2020;125:38–45. <https://doi.org/10.1016/j.ejca.2019.10.027>.
 51. Eggermont AMM, Bellomo D, Arias-Mejias SM, et al. Identification of stage I/IIA melanoma patients at high risk for disease relapse using a clinicopathologic and gene expression model. *Eur J Cancer*. 2020;140:11–8. <https://doi.org/10.1016/j.ejca.2020.08.029>.
 52. Chen J, Xu Y, Zhou Y, Wang Y, Zhu H, Shi Y. Prognostic role of sentinel lymph node biopsy for patients with cutaneous melanoma: A retrospective study of surveillance, epidemiology, and end-result population-based data. *Oncotarget*. 2016;7(29):45671–7. <https://doi.org/10.18632/oncotarget.10140>. PMID:27344178;PMCID:PMC5216751.
 53. Aiken TJ, Stahl CC, Schwartz PB, et al. Sentinel lymph node biopsy is associated with increased cost

- in higher risk thin melanoma. *J Surg Oncol*. 2021;123(1):104–9. <https://doi.org/10.1002/jso.26225>.
54. Morton DL, Cochran AJ, Thompson JF, et al. Sentinel node biopsy for early-stage melanoma: accuracy and morbidity in MSLT-I, an international multicenter trial. *Ann Surg*. 2005;242(3):302–11; discussion 311–3. <https://doi.org/10.1097/01.sla.0000181092.50141.fa>.
 55. Whitman ED, Koshenkov VP, Gastman BR. Integrating 31-gene expression profiling with clinicopathologic features to optimize cutaneous melanoma sentinel lymph node metastasis prediction. *JCO Precision Oncology*. 2021;1466–79. <https://doi.org/10.1200/po.21.00162>
 56. Meves A, Nikolova E, Heim JB, et al. Tumor cell adhesion as a risk factor for sentinel lymph node metastasis in primary cutaneous melanoma. *J Clin Oncol*. 2015;10;33(23):2509–15. <https://doi.org/10.1200/JCO.2014.60.7002>.
 57. Mulder EEAP, Dwarkasing JT, Tempel D, et al. Validation of a clinicopathological and gene expression profile model for sentinel lymph node metastasis in primary cutaneous melanoma. *Br J Dermatol*. 2021;184(5):944–51. <https://doi.org/10.1111/bjd.19499>
 58. 2020 Clinically Tested Patients, Data on File from Castle Biosciences Laboratory Management System; Castle Biosciences Inc., 07.29.2021.
 59. Marson, J., Litchman, G., Svoboda R, et al. Assessment of the 31-gene expression profile test by dermatologists: a cross-sectional survey from national dermatology conferences. *SKIN The Journal of Cutaneous Medicine*. 2021;5(2):101–7. <https://doi.org/10.25251/skin.5.2.4>.
 60. Dillon LD, Gadzia JE, Davidson RS, et al. Prospective, Multicenter clinical impact evaluation of a 31-gene expression profile test for management of melanoma patients. *SKIN The Journal of Cutaneous Medicine*. 2018;2(2):111–21. <https://doi.org/10.25251/skin.2.2.3>
 61. Schuitevoerder D, Heath M, Cook RW, et al. Impact of gene expression profiling on decision-making in clinically node negative melanoma patients after surgical staging. *J Drugs Dermatol*. 2018;17(2):196–9.
 62. Berger AC, Davidson RS, Poitras JK, et al. Clinical impact of a 31-gene expression profile test for cutaneous melanoma in 156 prospectively and consecutively tested patients. *Curr Med Res Opin*. 2016;32(9):1599–604. <https://doi.org/10.1080/03007995.2016.1192997>.
 63. Farberg AS, Glazer AM, White R, Rigel DS. Impact of a 31-gene expression profiling test for cutaneous melanoma on dermatologists' clinical management decisions. *J Drugs Dermatol*. 2017;16(5):428–31.
 64. Hyams DM, Covington KR, Johnson CE, Plasseraud KM, Cook RW. Integrating the melanoma 31-gene expression profile test with surgical oncology practice within national guideline and staging recommendations. *Future Oncol*. 2021;17(5):517–27. <https://doi.org/10.2217/fon-2020-0827>.
 65. Scott AM, Dale PS, Conforti A, Gibbs JN. Integration of a 31-gene expression profile into clinical decision-making in the treatment of cutaneous melanoma. *Am Surg*. 2020;86(11):1561–4. <https://doi.org/10.1177/0003134820939944>.
 66. Mirsky R, Prado G, Svoboda R, Glazer A, Rigel D. Management decisions made by physician assistants and nurse practitioners in cutaneous malignant melanoma patients: impact of a 31-gene expression profile test. *J Drugs Dermatol*. 2018;17(11):1220–3.
 67. Litchman GH, Prado G, Teplitz RW, Rigel D (2020). A systematic review and meta-analysis of gene expression profiling for primary cutaneous melanoma prognosis. *SKIN The Journal of Cutaneous Medicine*.4(3):221–37. <https://doi.org/10.25251/skin.4.3.3>
 68. Hariton E, Locascio JJ. Randomised controlled trials—the gold standard for effectiveness research: Study design: randomised controlled trials. *BJOG*. 2018;125(13):1716. <https://doi.org/10.1111/1471-0528.15199>.
 69. Paik S, Shak S, Tang G, et al. A multigene assay to predict recurrence of tamoxifen-treated, node-negative breast cancer. *N Engl J Med*. 2004;351(27):2817–26. <https://doi.org/10.1056/NEJMoa041588>.
 70. Paik S, Tang G, Shak S, et al. Gene expression and benefit of chemotherapy in women with node-negative, estrogen receptor-positive breast cancer. *J Clin Oncol*. 2006;24(23):3726–34. <https://doi.org/10.1200/JCO.2005.04.7985>